PEPTIDES CONTAINING β-Turns I--<u>CYCLO</u>-(GLY-L-CYS-GLY)<sub>3</sub> TRIPLY BRIDGED BY 1,3,5-<u>TRIS</u>-(THIOMETHYL)BENZENE

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Abstract: Reaction of cyclo-(Gly-L-Cys-Gly)<sub>3</sub> with 1,3,5-tris-bromomethylbenzene yields a tris thioether of the cyclic nonapeptide in 28% yield. Both <sup>1</sup>H and <sup>1</sup>3C NMR spectra are consistent with a molecule of 3-fold symmetry; the temperature dependences of chemical shifts of the amide hydrogens are consistent with a structure composed of three  $\beta$ -turns.

Although the  $\beta$ -turn<sup>1</sup> attracted interest much later than the  $\alpha$ -helix and the  $\beta$ -sheet, it appears to be at least as prevalent in protein structures<sup>2</sup> and accumulating evidence suggests that it may be assumed by many peptide hormones in the hormone-receptor complex.<sup>3</sup> Recently Chou and Fasman have reported a simple analysis for predicting the probability that a given sequence of four amino acids will assume a  $\beta$ -turn conformation in a protein structure.<sup>4</sup> The  $\beta$ -turn or certain closely related structures partially define the orientation of a number of cage-type ionophores such as valinomycin.<sup>5</sup>



As seen in 1, the  $\beta$ -turn is a relatively rigid structure that is defined by only four amino acid residues. In principle rigid subunits can be linked to form larger arrays with defined structures, and as part of a general program directed toward unusual cage-type structures that can be formed from polypeptide precursors, we were led to ask, what simple structures might be formed by linkages of subunits consisting of tetrapeptides that assume the  $\beta$ -turn conformations. The simplest of these is a cyclic hexapeptide consisting of two  $\beta$ -turns, 2.<sup>6</sup> One example of cyclic nonapeptide 3 that contains three  $\beta$ -turns, has been reported,<sup>7</sup> in which R is a leucine side chain.



In this paper we report synthesis of a <u>cyclo</u>nonapeptide  $(Gly-\underline{L}-Cys-Gly)_3$  as its tri S-benzyl derivative, which was prepared by the reaction sequence of Scheme 1, using 2ethyl-7-hydroxybenzisoxazolium fluoroborate<sup>7</sup>  $\underline{A}$  and 2-ethylbenzisoxazolium fluoroborate<sup>8</sup> 5 as the amide-forming reagents. The cyclization of the active ester of the linear nonapeptide 9 is noteworthy for the high yield that was observed. (Structures marked with asterisks were characterized by TLC, HPLC, <sup>1</sup>H NMR, and satisfactory elemental analysis.)



The 250 MHz <sup>1</sup>H NMR spectrum of 10 at 25°C in DMS0<sup>-D6</sup> showed three amide resonances at 7.99 $\delta$ (t,3H), 8.18 $\delta$ (d,3H), and 8.73 $\delta$ (t,3H). The temperature dependences of these resonances are 2.4, 5.0, and 8.0 x 10<sup>-3</sup> ppm/°C, respectively. Temperature dependences of less than 3 x 10<sup>-3</sup> ppm/°C are characteristic of internally hydrogen bonded amide NH groups, and

values of greater than 6 x  $10^{-3}$  ppm/°C are expected for amide NH groups that are hydrogen bonded intermolecularly with solvent.<sup>10</sup> Since the triplet resonances must be assigned to NH residues of glycines, we interpret the NMR evidence as consistent with structure 3 in which glycine residues appear at site 4 of the turn structure 1. Models suggest that bent hydrogen bonds can exist (dashed lines) in a stable conformation of 3. Taken together with the intermediary value seen for the temperature dependence of the cysteine NH supports structure 3 with R = CH<sub>2</sub>-S-Bz1 for the peptide 10.



Structure ]] is a more rigid analog of 3 in which cysteine residues appear at  $\beta$ -turn sites 1 and 4 (structure ]), and the sulfurs are bound as thioethers by a 1,3,5-tris-(methylene)benzene residue. When 10 was reduced with sodium in liquid ammonia and then caused to react with 1,3,5-tris(bromomethy1)benzene, a product was isolated by preparative HPLC (Whatman Magnum 9 ODS-2 C-18 reverse phase column; methanol-water eluant) which was characterized having the molecular formula of  $\car{ll}$  by elemental analysis  $\car{ll}$  and by field desorption MS  $(M^+, 766)$ . Although ]] is an insoluble substance that has thus far defied attempts at macro crystallization, important aspects of its structure can be deduced from its spectroscopic behavior. At 62.8 MHz, the <sup>13</sup>C NMR spectrum of ]] in DMSO-d<sup>6</sup> exhibits resonances at 6 172.6(Cys C=0,3C), 169.8(Gly C=0,3C), 169.3(Gly C=0,3C), 136.9(Ar, 3C), 128.9 (Ar, 3C) and 52.5 (Cys  $\alpha$ -C, 3C). The remaining four carbon resonances are obscured by solvent. The amide NH resonances at 250 MHz,  $25^{\circ}$ C, DMSO D<sup>6</sup>, appear at  $8.95\delta(t, 3H)$ ,  $7.82\delta(t, 3H)$ , and  $7.29\delta(d, 3H)$ , with respective temperature dependencies of -4.5,-1.4, and -1.7 ppb/°C. These results are consistent with a structure with three-fold symmetry in which the NH groups of the cysteine and one glycine residue in each Gly-Cys-Gly subunit are involved in intramolecular hydrogen bonds. The available data are thus consistent with structure ]] for this substance.

A Chou-Fasman analysis<sup>4</sup> of the probability of  $\beta$ -turn formation by the sequences Cys-Gly-Gly-Cys (as in 11) and Gly-Cys-Gly-Gly (as in 3) give values of 3.1 x 10<sup>-4</sup> and 1.6 x 10<sup>-4</sup>, respectively. Since turns are predicted by this model for values larger than 7.5 x 10<sup>-5</sup>, both structures are predicted to have turn conformations. Space-filling models for 11 suggest a rigid flowerpot-like structure that may exhibit interesting binding properties. Syntheses and study of analogs of 11 that should exhibit more favorable solubility properties are in process and will be reported subsequently.

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## References:

- J. Crawford, W. Lipscomb and C. Schellman, <u>Proc. Natl. Acad. Sci.</u>, 70, 538 (1973);
  G.M.J. Schmidt, D.C. Hodgkin, and B.M. Oughton, <u>Biochem. J.</u>, 65, 752 (1957); R. Schwyzer and A. Tun-Kyi, <u>Helv. Chim. Acta</u>, 45, 859 (1962). For a recent review, see: J.A. Smith and L.G. Pease, <u>CRC Critical Reviews in Biochem</u>., 8, 315 (1980).
- P. Lewis, F. Momany, and H. Scheraga, <u>Proc. Nat. Acad. Sci US</u>, 68, 2293 (1979); P. Chou and G. Fasman, <u>J. Mol. Biol.</u>, 74, 263 (1973).
- 3. R. Nutt, D. Veber, and R. Saperstein, J. Am. Chem. Soc., 102, 6539 (1980).
- 4. P.Y. Chou and G.D. Fasman, Ann. Rev. Biochem., 47, 251 (1978).
- 5. Yu A. Ovchinnikov and V.T. Ivanov, Tetrahedron 30, 1871 (1974).
- 6. R. Schwyzer and U. Ludescher, Helv. Chim Acta, 52, 2033 (1969).
- K.D. Kopple and J. Savrda. In <u>Peptides 1971</u>, H. Nesvadba, ed., North-Holland, Amsterdam, (1973), p. 400.
- 8. D.S. Kemp et al., Tetrahedron 30, 3969 (1974).
- 9. D.S. Kemp, Tetrahedron 23, 2001 (1967).
- Yu A. Ovchinnikov and V.T. Ivanov, <u>Tetrahedron</u> 31, 2177 (1975); V.J. Hruby. In <u>Chemistry and Biochemistry of Amino Acids, Peptides and Proteins, Vol. 3</u>, B. Weinstein, ed., Marcel Dekker, New York, (1974), p. 1.
- 11. Calcd for  $C_{30}H_{39}O_{9}N_{9}S_{3}H_{2}O$ : C 45.96, H 5.27, N 16.08, S 12.27. Found: C 46.00, H 5.32, N 16.06, S 11.91.

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